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Efficient Synthesis of *lactoneo* Series Antigens H, Lewis X (Le^x), and Lewis Y (Le^y)¹

Rainer Windmüller and Richard R. Schmidt*

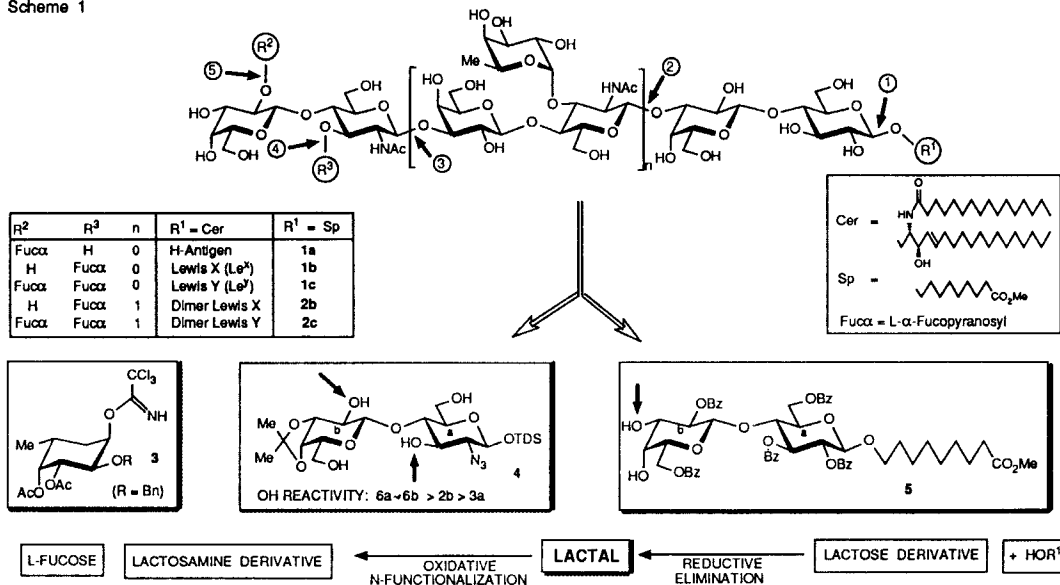
Fakultät Chemie, Universität Konstanz,

Postfach 5560 M 725, D-78434 Konstanz, Germany

Abstract: Regioselective 6a,6b-di-O-benzylation of azidolactose 4 furnished derivative 6 which permitted ensuing regioselective 2b-O-fucosylation, 2b,3a-di-O-fucosylation, or regioselective 2b-O-benzylation and then 3a-O-fucosylation, respectively, thus providing directly H, Le^y, and Le^x antigen building blocks 8a-c. The derived trichloroacetimidates 9a-c offered regio- and β-selective glycosylations of partially O-protected acceptors 5 and 13, affording spacer-linked H, Le^x, Le^y, dimer Le^x and Le^y intermediates 12a-c and 14b,c, respectively, which could be readily transformed into target molecules 1a-c and 2b,c. Thus, a most straightforward and efficient synthesis of this antigen series is exhibited.

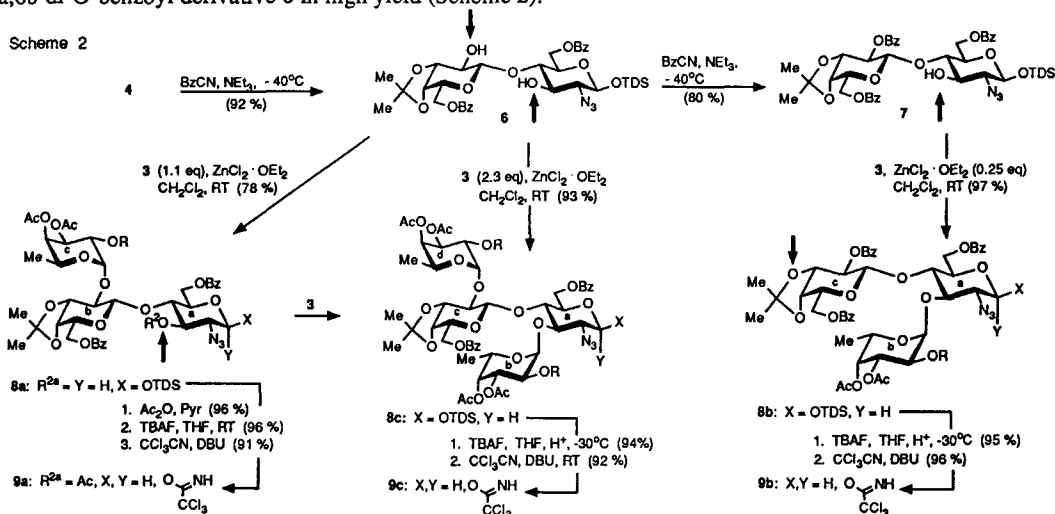
Prominent tumor associated antigens are the Lewis X (Le^x) and Lewis Y (Le^y) determinants²; these glycosphingolipids, including the important sialyl Le^x epitope³, are members of the *lactoneo* family, i.e. *lacto* series type 2 chain. Syntheses of Le^x and Le^y antigens have employed various strategies, various building blocks and/or protecting group patterns⁴⁻¹⁴. We would like to report here on a very efficient synthesis of spacer-linked antigens H, Le^x, Le^y, dimer Le^x, and dimer Le^y (Scheme 1, 1a-c, 2b,c) which is based on lactal derived azidolactose¹⁴⁻¹⁶ as outlined in the retrosynthetic strategy (Scheme 1); lactal is obtained from lactose *via reduct-*

Scheme 1



tive elimination. The retrosynthesis exhibits that starting from lactal, the sequence *oxidative azide introduction*, regioselective O-protection, anomeric carbon activation, glycoside bond formation, and deprotection is closely related to the sequence regioselective lactal protection, *oxidative nitrogen introduction*, anomeric carbon activation via aziridine formation and concomitant glycoside bond formation, and then deprotection, which has been reported recently¹⁰.

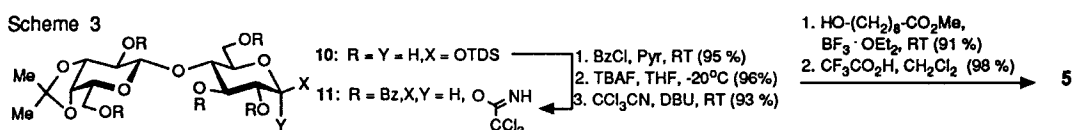
The decisive building blocks in our approach are O-(3,4-O-acetyl-2-O-benzyl-L-fucopyranosyl)-trichloroacetimidate (**3**), readily available from L-fucose¹⁵, hexyldimethylsilyl (TDS) 2a-azido-2a-deoxy-3b,4b-O-isopropylidene-lactoside **4**, readily available from lactal^{15,16}, and spacer-linked penta-O-benzoyl-lactoside **5**, readily available from lactose, as shown below. An important aspect is regioselective O-benzoylation of **4** which was found to exhibit an 6a – 6b > 2b > 3a reactivity order¹⁵. Thus, regioselective O-benzoylation of **4** with 2.2 equivalents of benzoyl cyanide in the presence of triethylamine at -40°C afforded 6a,6b-di-O-benzoyl derivative **6** in high yield (Scheme 2).



6 can be directly fucosylated with 1.1 equivalents of **3** in the presence of ZnCl₂ · OEt₂ as the catalyst to afford regioselectively 2b-O-fucosyl derivative **8a** required for H antigen construction; however, when 2.3 equivalents of **3** are employed in this reaction, practically exclusively **8c** the 2b,3a-di-O-fucosyl derivative of **6** is obtained, which is the building block for Le^y antigen construction. The reactivity difference of 2b-OH over 3a-OH in **6** can be also employed for the regioselective 2b-O-benzoylation. Thus, with an additional equivalent of benzoyl cyanide/triethylamine at -40°C 2b,6a,6b-tri-O-benzoyl derivative **7** is obtained in high yield; then fucosylation with **3** in the presence of ZnCl₂ · OEt₂ as the catalyst (0.25 equivalents) afforded trisaccharide **8b**, constituting an Le^x antigen building block. Intermediates **8a-c** were readily transformed into the corresponding trichloroacetimidates **9a-c** required as glycosyl donors.

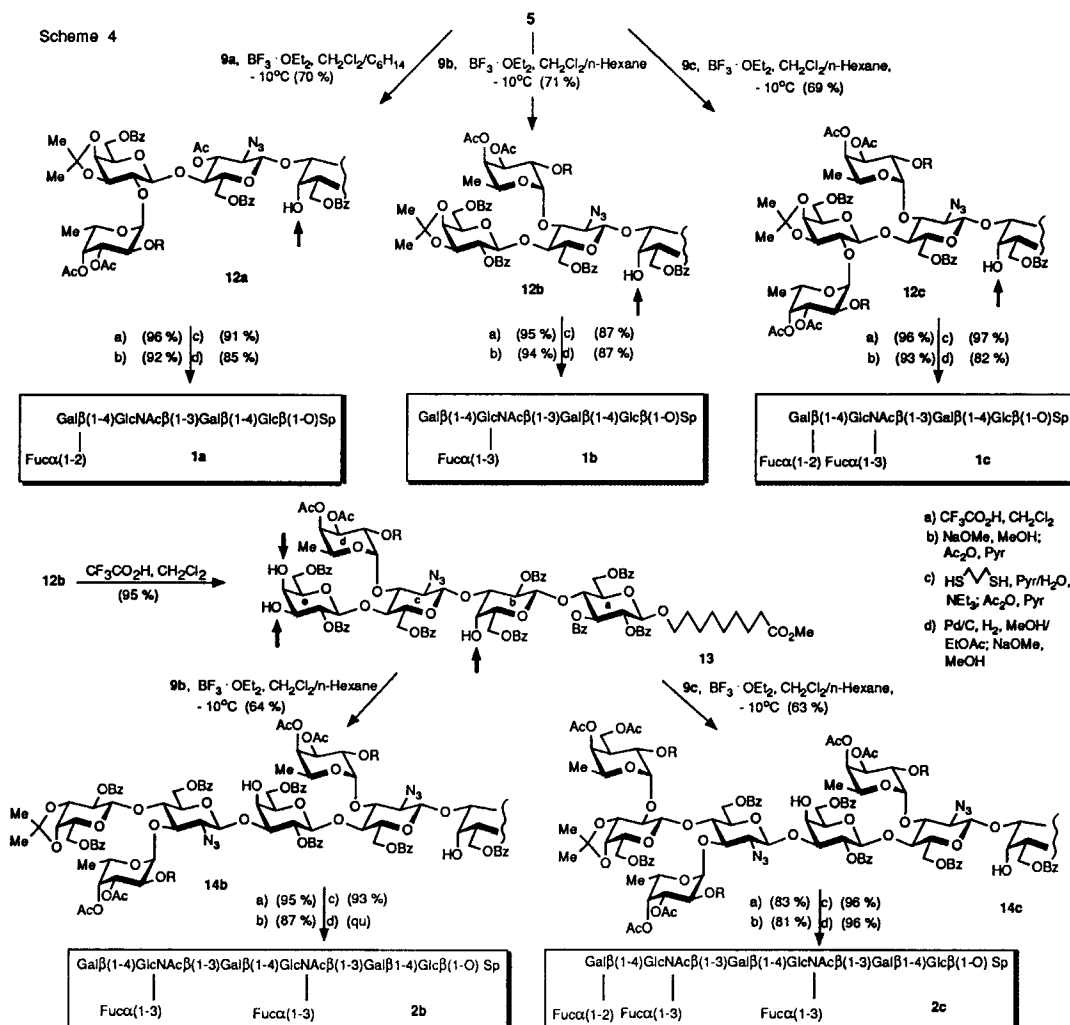
Spacer-linked lactose building block **5** was obtained from lactose derivative **10**¹⁵ via per-O-benzoylation, TDS group removal with tetra-*n*-butylammonium fluoride (TBAF), and trichloroacetimidate formation with CCl₃CN/DBU (→ **11**, Scheme 3). Glycosylation of the Lemieux spacer with **11** in the presence of BF₃ · OEt₂ as the catalyst and then de-O-isopropylidenation afforded **5** in very high overall yield.

Scheme 3



The combination of the building blocks of Schemes 2 and 3 for obtaining the target molecules turned out to be very straightforward. Reaction of glycosyl donors **9a-c** with acceptor **5** in CH₂Cl₂/*n*-hexane at -10°C in the presence of BF₃·OEt₂ as the catalyst afforded the H, X, and Y antigen intermediates **12a-c** in good yields, thus exhibiting the expected regioselective reaction at 3-OH of the galactose moiety of **5** (Scheme 4). Intermediates **12a-c** were transformed by the same highyielding four step sequence (consisting of acid catalyzed isopropylidene group removal, deacylation and per-O-acetylation, azide group reduction and N-acetylation, and hydrogenolytic debenzoylation and O-deacetylation) into the desired target molecules **1a-c**.¹⁷

Scheme 4



The building blocks **9b,c** and **12b** could be also successfully employed for the synthesis of spacer-linked dimer Le^x and Le^y antigens **2b,c**. Removal of the O-isopropylidene group in **12b** with CF_3CO_2H/CH_2Cl_2 afforded **3e**, **4b**, **4e**-O-unprotected derivative **13** in quantitative yield. Glycosylation of **13** with donors **9b** and **9c** under the above described conditions furnished under exclusive **3e**-O-attack the protected dimer Le^x and Le^y intermediates **14b,c**, respectively, again in acceptable yields; the same sequence of reactions as described for **12a-c** transformed **14b,c** into target molecules **2b,c**¹⁷ in high overall yield.

In summary, diastereoselective lactal transformation into silyl **3b,4b**-O-isopropylidene-2-azidolactoside **4** provides a stereochemically secured starting material which permits highly regioselective reactions at the remaining hydroxy groups, thus furnishing via simple benzoylations and highyielding fucosylations directly the desired H, Le^x , and Le^y antigen building blocks. Their transformation into trichloroacetimidates affords reactive glycosyl donors for regioselective β -glycosylations of the readily accessible partially O-protected acceptors **5** and **13**, thus providing the protected target molecules in very few steps. For deprotection, highyielding standard procedures can be employed.

References and Notes

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17. Values of $[\alpha]_D$ and δ_H : **1a**: $[\alpha]^{20} = -30.0$ (c = 0.5, H_2O) 1H -NMR (250 MHz, D_2O): $\delta = 4.23$ (d, $J_{1,2} = 8.7$ Hz, 1 H, 1-H), 4.28 (d, $J_{1,2} = 8.3$ Hz, 1 H, 1-H), 4.35 (d, $J_{1,2} = 7.6$ Hz, 1 H, 1-H), 4.50 (d, $J_{1,2} = 8.2$ Hz, 1 H, 1c-H), 5.11 (bs, $J_{1,2} < 1$ Hz, 1 H, 1e-H). **1b**: $[\alpha]^{20} = -36.0$ (c = 1, H_2O) 1H -NMR (250 MHz, D_2O): $\delta = 4.24$ (d, $J_{1,2} = 7.9$ Hz, 1 H, 1-H), 4.27 (d, $J_{1,2} = 7.7$ Hz, 1 H, 1-H), 4.28 (d, $J_{1,2} = 8.1$ Hz, 1 H, 1-H), 4.51 (d, $J_{1,2} = 8.0$ Hz, 1c-H), 4.93 (d, $J_{1,2} = 3.9$ Hz, 1 H, 1d-H). **1c**: $[\alpha]^{20} = -71.0$ (c = 1, H_2O) 1H -NMR (250 MHz, D_2O): $\delta = 4.23$ -4.40 (3 d, $J_{1,2} = 8.8$ Hz, $J_{1,2} = 8.4$ Hz, $J_{1,2} = 8.7$ Hz, 3 H, 1a-H, 1b-H, 1e-H), 4.53 (d, $J_{1,2} = 8.2$ Hz, 1 H, 1c-H), 4.93 (d, $J_{1,2} = 3.9$ Hz, 1 H, 1d-H), 5.09 (d, $J_{1,2} = 2.3$ Hz, 1 H, 1f-H). **2b**: $[\alpha]^{20} = -56$ (c = 1, H_2O) 1H -NMR (250 MHz, D_2O): $\delta = 4.25$ -4.36 (4 d, $J_{1,2} = 7.6$ Hz, $J_{1,2} = 8.3$ Hz, $J_{1,2} = 8.2$ Hz, $J_{1,2} = 8.0$ Hz, 4 H, 1a-H, 1b-H, 1e-H, 1h-H), 4.54 (2 d, $J_{1,2} = 8.5$ Hz, 2 H, 1c-H, 1f-H), 4.95 (d, $J_{1,2} = 4.0$ Hz, 1 H, 1d-H), 4.97 (d, $J_{1,2} = 4.1$ Hz, 1 H, 1g-H). **2c**: $[\alpha]^{20} = -68$ (c = 0.5, H_2O) 1H -NMR (250 MHz, D_2O): $\delta = 4.25$ -4.36 (4 d, $J_{1,2} = 8.2$ Hz, $J_{1,2} = 8.4$ Hz, $J_{1,2} = 8.0$ Hz, $J_{1,2} = 7.9$ Hz, 1a-H, 1b-H, 1e-H, 1h-H), 4.55 (2 d, $J_{1,2} = 7.9$ Hz, 2 H, 1c-H, 1f-H), 4.95 (bs, 2 H, 1d-H, 1g-H), 5.11 (bs, 1 H, 1i-H).